

Specification

RNase A

A2760

Physical Description:	Solid
Product Code:	A2760
Product Name:	RNase A
Short Description:	delivery form: salt-free, freeze dried
Specifications:	Assay: approx. 70 % Activity: min. 70 U/mg (Kunitz)
WGK:	1
Storage:	-20°C
Origin:	from bovine pancreas
M:	~13700 g/mol
CAS:	9001-99-4
EINECS:	232-646-6
CS:	35079090

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Comment

Ribonuclease A (RNase A) is an endoribonuclease, that specifically cleaves single-stranded RNA 3' to pyrimidine residues (cytosine, uracil). Thereby, it generates pyrimidine-3'-phosphate or oligonucleotides with terminal pyrimidine-3'-phosphates. The pH-optimum is in the range of 7.0 - 7.5. RNase A is used for the purification of RNA-free DNA, for the removal of non-hybridized regions of RNA \: DNA-hybrides or as a molecular weight marker. The enzyme is inhibited by diethyl pyrocarbonate (DEPC), guanidinium salts (4 M GuaSCN), β -mercaptoethanol, heavy metals, vanadyl-ribonucleoside-complexes, RNase-inhibitor from human placenta and competitively by DNA, respectively. Regarding the latter, the effect of denatured DNA is higher than by native nucleic acids. Nevertheless, RNase A is very active under very different conditions and difficult to inactivate. At low salt-concentrations (up to 100 mM NaCl), RNase A cleaves single- and double-stranded RNA and RNA in RNA \: DNA- hybrides. Under high salt concentrations (>300 mM NaCl) single-stranded RNA is cleaved only. To remove the enzyme from samples, it has to be digested by proteinase K (frequently, SDS at a final concentration of 0.6 % is added) and several phenol extractions are required. (Applications\: Enzymatic manipulation of DNA and RNA\; ref. 1 Suppl. 8 p. 3.13.1; minipreps of plasmid-DNA\; ref. 1 Suppl. 24 p. 1.6.6; *inSitu*-hybridisation of cellular RNA\; ref. 1 Suppl. 7 p. 14.3.8; removal of RNA from plasmid preparations\; ref. 2 p. 1.51) Stock solutions are prepared at concentrations from 1 - 10 mg/ml in 10 mM Tris \cdot HCl, pH 7.5; 15 mM NaCl or in 10 mM Tris \cdot HCl, pH 7.5; 1 mM EDTA, pH 8.0 (TE buffer). The recommended working concentration is 10 μ g/ml (removal of RNA from plasmid preparations; 1 hr, RT) or 100 ng/ml (preparation of "blunt ends" of double-stranded cDNA). **Unit-definition\:** One unit of activity is defined as that amount of enzyme which causes the hydrolysis of RNA to yield a velocity constant, $k = 1$, at 25°C and pH 5.0 (Kunitz-Unit). **Inactivation of DNase activity\:** A protocol (ref. 2) suggests to dissolve 10 mg/ml RNase A in 0,01 M Sodium acetate (pH 5,2), to heat to 100°C for 15 minutes in a water bath and to cool down to room temperature very slowly. The pH value is equilibrated by adding 0.1-fold the volume of 1 M Tris-Cl (pH 7,4). **Caution\:** Heating solutions of RNase A to inactivate DNase may not be satisfactory since RNase activity may be lost if precipitate formation occurs. For applications that require DNase-free RNase A we recommend our product A3832, RNase A (DNase-free). **Stability\:** RNase A aggregates during lyophilizing and storage. It has a high affinity to glass surfaces, which has to be taken into consideration. At neutral pH (e. g. in PBS pH 7.4) and high concentrations (> 10 mg/ml) the enzyme will precipitate. At +4°C (lyophilized) it is stable for several years (dry storage), in solution (-20°C) several years or (+4°C) several weeks.

Bibliography

(1) Ausubel, F.A., Brent, R., Kingston, R.E., Moore, D.D., Seidman, J.G., Smith, J.A. & Struhl, K. (eds.) (1995) *Current Protocols in Molecular Biology*. Page 3.13.1 Suppl. 8; Greene Publishing & Wiley-Interscience, New York (2) Sambrook, J. & Russel, D.W. (2001) *Molecular Cloning\:* A Laboratory Manual, 3rd Edition Page A4.39. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York. (3) Melton, D.A. et al. (1984) *Nucleic Acids Res.* **12**, 7035-7056 Efficient *in vitro* synthesis of biologically active RNA and RNA hybridization probes from plasmids containing the SP6 promoter. (4) Winter, E. et al. (1985) *Proc. Natl. Acad. Sci. USA* **82**, 7575-7579 A method to detect and characterize point mutations in transcribed genes.